



# **The Nexus between Polyphenols and Gut Microbiota and Their Interplay in Human Health: A Brief Review**

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# **Abstract**

Polyphenols are a broad class of naturally occurring substances in plants and have drawn extensive attention as they may possess promising health-promoting benefits. Recently, gut microbiota and polyphenol interactions have been directly linked to the well-being of humans. The classification, sources, and interactions of polyphenols with the gut microbiota are presented in this review, highlighting their key health benefits in humans. Polyphenols undergo complex transformations within the gastrointestinal tract and interact with the gut microbiota, a varied collection of bacteria living in the digestive system. The interactions substantially influence the composition, functioning, metabolic activity, and gut microbiota diversity. Research indicates that polyphenols may possess prebiotic-like properties, favouring *Lactobacilli* and *Bifidobacteria* growth, among other beneficial bacteria. The fermentation of polyphenols is aided by these bacteria, which produce bioactive metabolites that may improve human health and well-being in various ways. Moreover, the alteration of gut microbiology caused by polyphenols has been linked to improvements in several health outcomes, including enhanced metabolic health, fortified immunological function, and a decreased susceptibility to chronic conditions like heart disease and certain forms of cancer. In summary, the intriguing relationship between polyphenols and gut microbiota has significant health implications for humans. Understanding these relationships can open the door to tailored dietary treatments and the development of functional foods to support a balanced gut microbiota and general well-being.

**Keywords:** Dietary Intake, Functional Food, Gut Microbes, Human Health, Polyphenols, Phytochemicals

# **1. Introduction**

Polyphenols are naturally occurring organic compounds that consist of several units of phenols. Plant-derived secondary metabolites are crucial in several applications, including therapeutic and industrial fields<sup>[1](#page-8-0)-[3](#page-8-0)</sup>. Polyphenols, predominantly derived from plants, represent one of the most extensively researched phytochemical classes. Several of these naturally occurring phytochemicals throughout the entire kingdom of plants make their way into the human diet through various fruits, vegetables, legumes, dry fruits, cereals, herbs and spices, beverages, and other food items<sup>[4-6](#page-8-0)</sup>. Whole plant foods contain over 8000 polyphenolic compounds, encompassing phenolic acids, flavonoids, lignans and stilbenes $^4\!\!$  $^4\!\!$  $^4\!\!$ .

Polyphenols are structurally complex, and the fundamental monomer is the phenolic ring. These compounds are categorized into phenolic alcohols and phenolic acids<sup>[7](#page-8-0)</sup>. Plant tissues contain a variety of polyphenols, which exist in different structural forms. These polyphenols are often present as glycosides or complex polymerized molecules with high molecular weights. These may include tannins or combined with various organic acids<sup>[7](#page-8-0)</sup>. Polyphenols, characterized by their large molecular weight of nearly 800 Daltons,

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can transversely cell membranes, allowing them to reach intracellular spaces where they function as phytochemicals or pigments<sup>[8](#page-8-0)</sup>.

The healthcare sector is becoming increasingly interested in polyphenols. Numerous research has showcased the health-promoting properties of polyphenols, highlighting their pivotal function in regulating metabolism, addressing chronic disorders, managing weight, and controlling cell proliferation. As many polyphenolic compounds are identified, their comprehensive understanding of long-term and shortterm health effects is still incomplete. Evidence from epidemiological tests, human trials, and animal studies suggests that several polyphenols have anti-inflammatory and antioxidant properties. These qualities might have preventive and/or therapeutic implications in various non-communicable disorders, like obesity, cardiovascular diseases, neurodegenerative and cancer $9-14$  $9-14$ .

## **2. Polyphenols Classification**

Polyphenols are categorized into various groups, encompassing phenolic acids (such as hydroxycinnamic and hydroxybenzoic acids), flavonoids (including flavonols, flavones, isoflavones, anthocyanins, and flavanones), stilbenes (like piceatannol and resveratrol), lignans (such as pinoresinol, sesamol, enterodiol, and sinol), and others including tannins





(non-hydrolyzable, and condensed tannins), xanthones, lignins, anthraquinones and chromones<sup>[1,8](#page-8-0)[,15](#page-9-0)</sup>. Figure 1 represents the classification of polyphenols.

#### **2.1 Phenolic Acids**

Nearly one-third of the total known polyphenols are represented by phenolic acids. The classification of phenolic acids is represented in Figure 2. The two subcategories of phenolic acids are hydroxycinnamic acids and hydroxybenzoic acid (Figure 3). In nature, phenolic acids are usually found in glycosylated or ester derivative forms rather than in their free form $16$ . Hydroxybenzoic acids are benzoic acid  $(C_7H_6O_2)$  derivatives. The subcategory of hydroxybenzoic acids includes notable examples such as ellagic acid, benzoic acid, gallic acid, protocatechuic acid, salicylic acid and vanillic acid $1,17$  $1,17$ .

The hydroxybenzoic acids found in olive products have various advantages: anti-inflammatory, cardioprotective actions, and antioxidant $1,8,18,19$  $1,8,18,19$ . Derived from cinnamic acid, hydroxycinnamic acid is a member of the aromatic acid class  $(C6-C3)^{20}$ . Common hydroxycinnamic acid examples include caffeic acid, cinnamic acid, coumaric acid, sinapinic acid, and ferulic acid<sup>[8](#page-8-0)</sup>. Cinnamic acid serves as a fundamental structural and biological component constituting cell organelles.

#### **2.2 Stilbenes**

Stilbenes are chemical compounds with a condensed structure consisting of a core ethylene component and a single phenyl group (Figure 4). The phenyl group is one of the terminal groups of carbon double bonds $2^1$ .





Stilbenes, which are phenolics of low molecular weight, are induced (phytoalexins) in response to both abiotic and biotic stressors $22$ . Common sources of stilbenes include peanut (Fabaceae), sorghum (Poaceae), pine (Pinaceae) and grape (Vitaceae) $^{23}$ . Resveratrol, one of the extensively studied and renowned stilbenes, is linked to a broad spectrum of pharmacological attributes and is recognized for its numerous health-promoting effects $24-26$ .

## **2.3 Flavanoids**

Plants synthesize flavonoids, which have a benzoγ-pyrone structure, from phenylalanine, tyrosine, and malonate via the phenylpropanoid route $27,28$ . The basic building block of flavonoids is the flavan nucleus, which consists of 15 carbon atoms organized into three rings known as A, B, and C. Variations in oxidation and substitution levels of these rings (A, B, C) give rise to different subclasses of flavonoid (Figure 5). Flavonoids are present in nature as glycosides, aglycones, and methylated derivatives $28$ . Flavonoids are divided into six subclasses: anthocyanidins, flavones, flavonols, flavan-3-ols, isoflavones, and flavanones $8,15$  $8,15$ as presented in Table 1. The examples of the different subclasses of flavonoids are given in Table 1. Flavonoids are predominantly found in foods such as onions, berries, grapes, apples, tea and cocoa, showcasing numerous health benefits.

#### **2.4 Lignans**

Lignans are dimeric compounds formed by the condensation of two phenylpropanoid C6‐C3 units at the β and β' carbon atoms, and they possess the ability to form additional linkages through lactone, ether, or carbon bonds. Their chemical composition resembles that of  $1,4$ -diarylbutan<sup>30</sup>, and these compounds are synthesized through the shikimic acid biosynthesis pathway<sup>31</sup>. Lignans can be classified into two chief subclasses: neolignans and classical lignans. Phenylpropane dimers with a  $β$ -β' bond are known as classical lignans, and there are six primary subtypes of classical lignans, namely —dibenzylbutyrolactones, aryl tetralin/aryl naphthalenes, dibenzocyclooctadienes, dibenzyl butanes, 2,6-diarylfurofurans and substituted tetrahydrofurans<sup>32-34</sup>. Neolignans exhibit a more diverse range of structures compared to classical lignans. There are fifteen subtypes of neolignans,



**Figure 5.** Flavan nucleus.

each selected by nature as well as the linkage position between the phenylpropane units $31,32,35$ . The commonest subtypes of neolignans include alkyl aryl ethers, 8-10-bicyclo[3.2.1]octanes, 8-30-bicyclo[3.2.1] octanes, 1,4-benzodioxanes, benzofurans, biphenyls, biphenyl ethers and cyclobutanes. Lignans are secondary metabolites found in vascular plants, exhibiting extensive occurrence throughout the flora. These chemicals possess a diverse range of physiological properties that improve human health $36$ .

#### **2.5 Tannins**

Tannins are widespread natural phenolic compounds abundantly present in the plant kingdom. They are categorized primarily into condensed and hydrolysable types. Polyflavonoid tannins belong to the condensed class, these being less commonly hydrolysable. While ellagitannins and gallotannins fall into the hydrolysable category<sup>37</sup>. Condensed tannins contain flavonoids (flavan 3-ol or flavan 3, 4-diol) with no sugar core. In contrast, hydrolysable tannins compose gallic and ellagic acids with a sugar core, primarily glucose<sup>[38,39](#page-10-0)</sup>.





Hydrolyzable tannins are less frequently found in nature than condensed tannins among the many forms of tannins $40,41$  $40,41$  $40,41$ . As a result, condensed tannins account for more than 90% of all commercial tannins and command a dominant market share worldwide<sup>[42](#page-10-0),[43](#page-10-0)</sup>.

## **3. Major Dietary Sources of Polyphenols**

Polyphenols are bioactive compounds majorly found in various fruits, vegetables, herbs, seeds, and beverages namely wine, beer, tea, coffee, chocolate and fruit juice. Dry legumes and grains possess minor amounts of polyphenols $44$ . Generally, these chemicals safeguard against plant pathogens and environmental factors. These compounds contribute to the sensory attributes of foods by providing flavours, colours, and astringency. The major dietary sources of polyphenols are represented in Table 2. Currently, polyphenols are receiving immense scientific attention as they possess a wide range of health benefits for humans.

#### **Table 2.** Major dietary sources of polyphenols



## **4. Polyphenols' Impact on Gut Microbiota**

The human gastrointestinal system contains a vast and varied population of many bacteria, comprising billions of cells<sup>64</sup>. This gut microbiota functions like an organ, actively contributing to the breakdown and utilization of dietary constituents. It significantly impacts human well-being by generating both advantageous and detrimental metabolites, safeguarding against pathogens, regulating the immune system, and providing defence against various illnesses $65,66$  $65,66$ . The relevance of gut microbiota in maintaining the host health's physiological balance and affecting the onset of various ailments is well understood. These include neurological diseases, obesity, diabetes, and inflammatory bowel disorder $67$ .

Multiple factors influence the human gut microbiota: genetics, age, stress, medications, and diet, particularly reflecting long-term dietary habits $68$ . Polyphenols, crucial in host health, interact with host physiology and metabolism, playing roles in immune stimulation, oxidative stress regulation, and protection against pathogenic infections<sup>69</sup>. Ample evidence suggests that dietary polyphenols directly influence the gut microbiome, fostering the growth of beneficial microbial species while suppressing harmful ones $67$ . Dietary polyphenols have been observed to engage with human and animal gut microbiota, specifically interacting with *Bifidobacteria* and *Lactobacilli*. This interaction has shown a notable increase in butyrate production, contributing to the reduction of colitis and serving as a preventive measure against colitisassociated colorectal cancer. Simultaneously, it leads to a decrease in the presence of harmful microbial species<sup>[70](#page-11-0)</sup>. Polyphenols exhibit a dual effect in the gut by selectively impeding the development of harmful microbes. For instance, flavonoids found in red wine demonstrated a mild inhibitory effect on *Clostridium*, while anthocyanins, ellagic acid in raspberry juice, and grape polyphenols displayed inhibitory actions against *Ruminococcus* and *Clostridium histolyticum*, respectively<sup>[71-73](#page-11-0)</sup>. Conversely, polyphenols promote the proliferation of beneficial gut bacteria. Grape polyphenols, gingerol in ginger, sorghum polyphenols, and tannin in pomegranate were observed to stimulate the growth of *Bifidobacterium*[74-76.](#page-11-0) Tannin also contributes to the development of *Lactobacillus*, while grape polyphenols and gingerol support the proliferation of *Enterococci*. *Roseburia*, *Prevotella*, and Lactic acid bacteria are abundant when fructooligosaccharides and sorghum polyphenols are present $73,75,76$ .

Studies conducted in living organisms have highlighted the effectiveness of polyphenol supplementation in modifying the gut microbiota of animal models. These interventions have demonstrated an augmentation in beneficial microbial populations and a concurrent reduction in harmful microbes. For instance, Orso's research involved administering a diet rich in tannin extracted from chestnut shells to a zebrafish model of intestinal inflammation. This diet notably facilitated the growth of advantageous bacteria, specifically Enterobacteriaceae and *Pseudomonas*[77](#page-11-0). Moreover, specific polyphenols exhibit

selective inhibition against pathogenic bacteria. Research on polyphenols derived from *Smilax china* L. rhizome showcased a decreased relative abundance of *Desulfovibrionaceae*, *Lachnospiraceae*, and *Streptococcaceae*[78.](#page-11-0) Additionally, grape pomace polyphenols were observed to reduce potentially harmful bacteria in humans, including *Escherichia coli, Proteus, Salmonella*, *Shigella*, and *Yersinia*[79](#page-11-0). The combined administration of quercetin and resveratrol has demonstrated significant inhibition of specific microbial groups—*Desulfovibrionaceae*, *Acidaminococcaceae*, *Coriobacteriaceae*, *Bilophila*, and *Lachnospiraceae*—potentially linked to diet-induced obesity $80$ . In another study, blueberry polyphenols administered to ovariectomized rats led to an increase in *Bacteroides dorei* and *Lachnoclostridium* and a decrease in Rikenellaceae and *Eubacterium*<sup>81</sup>. Additionally, the importance of polyphenol supplementation is highlighted by consistent results from animal experiments conducted both in vivo and in vitro, particularly flavonoids and anthocyanins, in increasing the quantity of gut-protective microorganisms in humans, such as *Lactobacillus* and *Bifidobacterium*[82](#page-11-0)[,83](#page-12-0).

The prebiotic properties of polyphenols promote the growth of good bacteria, which restricts the availability of nutrients for pathogenic bacteria and functions as a powerful antibacterial agent against them $84$ . Research focusing on the benefits of flavonoids from *Theobroma cacao*, present in cocoa-based foods and drinks, shows that these flavonoids ameliorate insulin resistance, change how glucose is metabolized, enhance endothelial tissue function, and lower oxidative stress $85$ . Certain substances found in dark chocolate, such as epicatechin and catechin, have been shown to decrease the activity of α-glucosidase, which hinders the intestinal absorption of glucose $85,86$  $85,86$  $85,86$ .

In a recent investigation utilizing a mouse model of ulcerative colitis to explore the impact of gallic acid, it was observed that it positively mitigated dysbiosis within the gut microbiota. The intake of gallic acid reduced certain detrimental bacteria, such as *Firmicutes* and *Proteobacteria*, while enhancing the presence of the Lactobacillaceae and Prevotellaceae families<sup>[87-89](#page-12-0)</sup>. Gallic acid can reduce environmental stress in beagle puppies, which are thought to be a great model for researching the human microbiota because of their similarity to humans. This reduction in stress levels

aids in the restoration of intestinal health by increasing *Lactiplantibacillus* and *Faecalibaculum* and decreasing *Shigella*, *Clostridium*, and *Escherichia*. It is crucial to emphasize that environmental stress is responsible for the imbalance of gut microbiota, increased inflammatory reactions, and disruptions in the integrity of the intestinal barrier $90,91$ . Furthermore, women in good health, the gut flora was positively affected by certain polyphenols in orange juice, particularly hesperidin and naringin. These polyphenols had a beneficial effect on the microbiota's composition and metabolic activity in addition to raising important blood biochemical markers like insulin sensitivity, lowdensity lipoprotein cholesterol, and glucose levels. This was demonstrated by higher production of Short-Chain Fatty Acids (SCFA) and increased *Lactiplantibacillus* and *Bifidobacterium* spp. populations. These results imply that eating oranges helps improve the gut microbiota and the related metabolites it $^{92}$  $^{92}$  $^{92}$ .

Polyphenols possess strong antibacterial effects that hinder the proliferation of harmful bacterial species, preventing gut biofilms. Several polyphenols possess antibacterial properties that target harmful bacteria, such as *Salmonella* and *Helicobacter pylori*. These include quercetin, hydroxytyrosol, resveratrol, and phenolic acids. For example, resveratrol can reduce the *E. coli* population and lessen the effects of heat stress in broilers $93$ . Quercetin has demonstrated effectiveness against *Salmonella enteritidis* and can impact the expression of genes linked to inflammation  $94$ . Their antimicrobial activity is mediated by direct suppression of bacterial species, reduction of pathogenic bacteria's ability to adhere, or interruption of ionic fluxes at the cell membrane<sup>[95](#page-12-0)</sup>.

Furthermore, polyphenols function as a source of nutrients that promote the growth of populations of specific bacteria, including *Lactobacilli* and *Bifidobacteria*, by acting as prebiotic-like agents<sup>[96,97](#page-12-0)</sup>. Consuming different polyphenols is associated with changes in the composition of gut bacteria, namely boosting the growth of *Bacteroides*, which have more enzymes for breaking down glycans. Supplementing broiler diets with ellagic acid during heat stress has demonstrated noteworthy impacts on the gut microbiota, intestinal barrier function, and antioxidant system. Including ellagic acid in the diet increased Nrf2/HO-1 mRNA levels in the ileum. In addition, it

decreased the quantities of specific bacterial species (e.g. *Actinomyces*, *Ruminococcus torques*, *Rothia*, *Neisseria*, and *Lautropia*) in the cecum in a manner that depended on the dosage. Dietary ellagic acid supplementation seemed to improve the body's ability to produce antioxidants, strengthen the intestinal barrier, and reduce heat-related injuries—possibly by controlling the gut microbiota $91$ . Additional research is necessary to examine ellagic acid's impact on chickens' gut microbiome under heat-stress conditions.

The role of gut bacteria in atherosclerosis is increasingly attracting attention. The gut microbiota and faecal metabolites are significantly impacted by ferulic acid. Notably, there has been a significant drop in the occurrence of *Firmicutes*, Erysipelotrichaceae, and *Ileibacterium*. These bacteria have been shown to have a favourable relationship with blood lipid levels in animals with atherosclerosis. Ferulic acid has been shown to reduce atherosclerotic damage, which may be partially explained by its influence on gut microbiota and lipid metabolism via the AMPKα/SREBP1/ACC1 pathway<sup>[98](#page-12-0)</sup>. Vanillin has also demonstrated a strong ability to alleviate abnormalities of the gut microbiota linked to obesity, which show up as a reduction in alpha- and beta-diversity. Vanillin increased the variety of *Verrucomicrobiota* and *Bacteroidetes* phyla while decreasing the prevalence of the *Firmicutes* phylum. Notably, vanillin inhibited the growth of bacteria that produce lipopolysaccharide (LPS) from the genus *Bilophila* and H<sub>2</sub>S-producing bacteria from the genus *Desulfovibrio*. However, it is unclear exactly how vanillin is related to enhanced gut microbiota in the fight against obesity $99$ .

Berries have gained more attention recently because they contain phytochemicals that may benefit human health. Berries are abundant in polyphenols, including anthocyanins, flavonols, and flavonols, hydrolyzable and condensed tannins, phenolic acids, stilbenes, and lignans. Berries contain polyphenols that hold promise as bioactive compounds for addressing cancer and associated metabolic disorders by influencing the composition of the gut microbiota<sup>100[-102](#page-13-0)</sup>.

Recent studies have explored the potential of tea polyphenols in regulating circadian rhythms and improving sleep, mood and immunity through interactions with gut microbiota. Tea polyphenols can modulate the composition and function of intestinal

microbiota, which in turn influences host circadian rhythms and metabolic processes<sup>103-105</sup>.

Apple polyphenol extract intervention improved depression-like behaviours in mice fed a high-sucrose diet by reducing stress hormones, increasing an antiinflammatory cytokine, inhibiting inflammatory pathways, improving gut barrier function, and modulating the gut microbiome. Apple polyphenol extract has potential as a dietary intervention for ameliorating depression-like disorders induced by a sugary diet $106$ .

Gut microbiota dysbiosis is involved in developing and progressing polycystic ovary syndrome, leading to increased inflammation and metabolic dysfunction. Polyphenols, including anthocyanin, catechins, and resveratrol, can regulate gut microbes and alleviate chronic inflammation in women with polycystic ovary syndrome, providing a potential novel therapeutic strategy. Targeting gut microbiota dysbiosis and reducing chronic inflammation using polyphenols may assist in the development of new treatments for polycystic ovary syndrome $107$ .

Oat phenolic compounds can alleviate a range of metabolic syndromes in mice fed a high-fat diet, including weight gain, glucose intolerance, elevated lipid levels, oxidative stress, and adipocyte hypertrophy. Oat phenolic compounds can improve the imbalance in gut microbiota caused by a high-fat diet, increasing the abundance of *Bacteroidetes*, reducing the diversity of *Firmicutes*, and modulating the levels of specific gut bacteria<sup>108</sup>.

## **5. Unveiling the Complexity: Polyphenols, Gut Microbiota, and Personalized Health**

Among the nearly eight thousand different polyphenols that have been identified, every polyphenol possesses a structure and features that are unique to itself. Studying the specific influence of these diverse compounds on the gut microbiota is challenging. It is possible that a sizeable amount of the polyphenols that are consumed will not be absorbed in the small intestine, and as a result, they will arrive in the colon without being digested. This factor contributes to the complexity of the situation. The potential health benefits they could provide could be jeopardized by this circumstance.

The way in which the polyphenols are processed by the bacteria in the gut varies from person to person, and the composition of an individual's gut microbiota determines the metabolic products that are produced.

Polyphenols can affect the variety and composition of the microbiota in the gut. They can either stimulate the growth of those bacteria that are good to the gut, or they can exhibit antibacterial qualities that lower the overall diversity and stability of the microbiota. Due to the distinct microflora in their intestinal tracts, polyphenol consumption can elicit a wide range of responses from individuals. As gut microbiota may be diverse in each individual, personalised strategies may be necessary to maximize the health advantages of polyphenols. Low dosages of polyphenols may have the ability to induce prebiotic effects, but high amounts may have antimicrobial effects that disrupt the equilibrium of the gut microbiota. The influence of polyphenols on intestinal microflora appears to be dose-dependent.

Studies conducted over a short period might not fully capture the comprehensive composition and functionality of the intestinal microbiota, it is of the utmost importance to investigate the temporal effects of polyphenols on the gut microflora. To acquire a thorough understanding of the impacts of polyphenol consumption over the long term, it is necessary to do additional research into the potential temporal dynamics. An investigation into the concept of personalized nutrition based on an individual's gut microbial profile is essential. In this context, identifying the most beneficial polyphenols for individuals with specific gut microbiota compositions is appropriate.

Gut microbiota has emerged as a major component of personalized medicine, and numerous approaches have been developed to modify the gut microbiota composition for therapeutic purposes. These approaches include pre- and probiotic interventions, microbiota transplantation or the inoculation with synthetic gut microbiota $109$ . The host's gut microbiota can improve drug efficacy, and inappropriate or unwanted gut bacteria can inactivate a drug. Gut microbiota can impact the safety and efficacy of drugs by enzymatic modification of drug structure, alteration of drug availability, and changes in bioactivity or toxicity<sup>109</sup>. While our understanding of the gut microbiota's effect on drug

<span id="page-8-0"></span>efficacy is still early, recent studies have underscored its pivotal role. The gut microbiota significantly influences the transformation of pharmaceuticals, affecting their bioactivity, toxicity, and its lifetime within the body. Elucidating the connection of gut microbiota diversity and complexity to drug efficacy can pave the way for precision medicine, aid in toxicology risk assessments, and enhance drug research and development in both pre-clinical and clinical studies $110$ .

It is important to gain mechanistic insight into how gut microbiota influences diseases and individual responses towards dietary or pharmaceutical management. Modifying these microbial landscapes and their functions can contribute to the development and progression of a wide range of human diseases, such as metabolic disorders, neurological conditions, and cancer. Furthermore, these modifications can influence the effectiveness of various interventions in treating these diseases, including diets, exercise, and medications. Exploring the role of gut microbiota in diseases and drug responses is a pathway towards advancing precision and personalized medicine $11$ .

To gain a deeper understanding of the intricate interactions between polyphenols and the gut microbiota on a molecular level, it is necessary to use technological advances such as high-throughput sequencing and metabolomics. The complex relationship that exists between polyphenols, gut microbiota, and the health of humans is something that should be investigated further by scientists and medical practitioners. This may pave the way for future developments in tailored nutrition and preventative healthcare treatment methods.

# **6. Conclusion**

Polyphenols and gut microbiota have a dynamic mutual influence, which greatly affects the health benefits of polyphenols in human beings. A better understanding of the dynamic interactions between the polyphenols and the gut microbiota will open new possibilities for preventing illness, enhancing health, and customizing interventions. Finally, to conclude, finding out more about these links could offer the possibility of personalized dietary therapies, as well as functional foods that help to maintain a healthy gut microbiome and, therefore, health.

# **7. References**

- 1. Prabhu S, Molath A, Choksi H, Kumar S, Mehra R. Classifications of polyphenols and their potential application in human health and diseases. J. Physiol. Nutr. Phys. Educ. 2021; 6(1):293-301. https://doi.org/10.22271/ journalofsport.2021.v6.i1e.2236
- 2. Sharma P, Roy M, Roy B. A review on the influence of floral biology, pollination efficiency and conservation strategies of endangered medicinal plant, *Rauvolfia serpentina* (L.) Benth. ex Kurz. Ann Phytomed. 2022; 11(1):86-98. https:// doi.org/10.54085/ap.2022.11.1.9
- 3. Chellammal HSJ. Fruits that heal: Biomolecules and novel therapeutic agents. Ann Phytomed. 2022; 11(1):7-14. https://doi.org/10.54085/ap.2022.11.1.2
- 4. Pandey KB, Rizvi SI. Plant polyphenols as dietary antioxidants in human health and disease. Oxid Med Cell Longev. 2009; 2(5):270-8. https://doi.org/10.4161/ oxim.2.5.9498 PMid:20716914 PMCid: PMC2835915
- 5. Chauhan D, Kumar K, Kumar S, Kumar H. Effect of incorporation of oat flour on nutritional and organoleptic characteristics of bread and noodles. Curr. Res. Nutr Food Sci Jour. 2018; 6(1):148-56. https://doi.org/10.12944/ CRNFSJ.6.1.17
- 6. Cheynier V. Phenolic compounds: from plants to foods. Phytochem Rev. 2012; 11(2-3):153-77. https://doi. org/10.1007/s11101-012-9242-8
- 7. Mitra S, Tareq AM, Das R, Emran TB, Nainu F, Chakraborty AJ, *et al.* Polyphenols: First evidence in the synergism and bioactivities. Food Reviews International. 2022; 39(7):1-23. https://doi.org/10.1080/87559129.2022.2026376
- 8. Singla RK, Dubey AK, Garg A, Sharma RK, Fiorino M, Ameen SM, *et al.* Natural polyphenols: Chemical classification, definition of classes, subcategories, and structures. J AOAC Int. 2019; 102(5):1397-400. https://doi. org/10.5740/jaoacint.19-0133 PMid:31200785
- 9. Pérez-Jiménez J, Neveu V, Vos F, Scalbert A. Identification of the 100 richest dietary sources of polyphenols: an application of the Phenol-Explorer database. Eur J Clin Nutr. 2010; 64(S3):S112-20. https://doi.org/10.1038/ ejcn.2010.221 PMid:21045839
- 10. Singh A, Holvoet S, Mercenier A. Dietary polyphenols in the prevention and treatment of allergic diseases. Clin Exp Allergy. 2011; 41(10):1346-59.https://doi.org/10.1111/ j.1365-2222.2011.03773.x https://doi.org/10.1111/j.1365- 2222.2011.03773.x PMid:21623967
- 11. Lecour S, Lamont KT. Natural polyphenols and cardioprotection. Mini Rev Med Chem. 2011; 11(14):1191- 9. https://doi.org/10.2174/138955711804586766 PMid:22070680
- 12. Cory H, Passarelli S, Szeto J, Tamez M, Mattei J. The role of polyphenols in human health and food systems: A mini-review. Front Nutr. 2018; 5(87):370438. https://doi.

<span id="page-9-0"></span>org/10.3389/fnut.2018.00087 PMid:30298133 PMCid: PMC6160559

- 13. Mounika M, Hymavathi TV. Nutrient and phytonutrient quality of Nutri cereals incorporated flour mix suitable for diabetics. Ann Phytomed. 2021; 10(1):132-40. https://doi. org/10.21276/ap.2021.10.1.14
- 14. Sharma N, Sarwat M. Functional foods for better health and weight loss. Ann Phytomed. An International Journal. 2022; 11(2)114-21. https://doi.org/10.54085/ap.2022.11.2.12
- 15. Rasouli H, Farzaei MH, Khodarahmi R. Polyphenols and their benefits: A review. Int J Food Prop. 2017; 20(Sup2):1- 42. https://doi.org/10.1080/10942912.2017.1354017
- 16. Pandey, K.B., Rizvi, S. I. Plant Polyphenols in Healthcare and Aging. In: Al-Gubory, K., Laher, I, editors. Nutritional Antioxidant Therapies: Treatments and Perspectives. Cham. Springer; 2017. https://doi.org/10.1007/978-3-319-67625- 8\_11
- 17. Ozcan T, Akpinar-Bayizit A, Yilmaz-Ersan L, Delikanli B. Phenolics in human health. Int J Chem Eng Appl. 2014; 5(5):393-6. https://doi.org/10.7763/IJCEA.2014.V5.416
- 18. Shahidi F, Yeo J. Bioactivities of phenolics by focusing on suppression of chronic diseases: A review. Int J Mol Sci. 2018; 19(6):1573. https://doi.org/10.3390/ijms19061573 PMid:29799460 PMCid: PMC6032343
- 19. Gupta S, Bishnoi J, Kumar N, Kumar H, Nidheesh T. *Terminalia arjuna* (Roxb.) Wight andArn.: Competent source of bioactive components in functional food and drugs. Pharma Innov. 2018; 7(3):223-31.
- 20. Teixeira J, Gaspar A, Garrido EM, Garrido J, Borges F. Hydroxycinnamic acid antioxidants: An electrochemical overview. Biomed Res Int. 2013; 2013:1-11. https:// doi.org/10.1155/2013/251754 PMid:23956973 PMCid: PMC3730368
- 21. Chou YC, Ho CT, Pan MH. Stilbenes: Chemistry and molecular mechanisms of anti-obesity. Curr Pharmacol Rep. 2018; 4(3):202-9. https://doi.org/10.1007/s40495-018- 0134-5
- 22. Bavaresco, L., Fregoni, C. Physiological Role and Molecular Aspects of Grapevine Stilbenic Compounds. In: Roubelakis-Angelakis, K.A. editors. Molecular Biology & Biotechnology of the Grapevine. Dordrecht. Springer; 2001. https://doi. org/10.1007/978-94-017-2308-4\_6
- 23. Dubrovina AS, Kiselev KV. Regulation of stilbene biosynthesis in plants. Planta. 2017; 246(4):597-623. https:// doi.org/10.1007/s00425-017-2730-8 PMid:28685295
- 24. Thomasset SC, Berry DP, Garcea G, Marczylo T, Steward WP, Gescher AJ. Dietary polyphenolic phytochemicals-promising cancer chemopreventive agents in humans? A review of their clinical properties. Int J Cancer. 2007; 120(3):451-8. https:// doi.org/10.1002/ijc.22419 PMid:17131309
- 25. Szkudelska K, Szkudelski T. Resveratrol, obesity and diabetes. Eur J Pharmacol. 2010; 635(1-3):1-8. https://doi. org/10.1016/j.ejphar.2010.02.054 PMid:20303945
- 26. Petrovski G, Gurusamy N, Das DK. Resveratrol in cardiovascular health and disease. Ann N Y Acad Sci. 2011; 1215(1):22-33. https://doi.org/10.1111/j.1749- 6632.2010.05843.x PMid:21261638
- 27. Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: An overview. ScientificWorldJournal. 2013; 2013(162750):1-16. https://doi.org/10.1155/2013/162750 PMid:24470791 PMCid: PMC3891543
- 28. Pietta PG. Flavonoids as antioxidants. J Nat Prod. 2000; 63(7):1035-42. https://doi.org/10.1021/np9904509 PMid:10924197
- 29. Guven H, Arici A, Simsek O. Flavonoids in our foods: A short review. J Basic Clin Health Sci. 2019; 3(2):96-106. https://doi.org/10.30621/jbachs.2019.555
- 30. Lewis N, Davin LB. Lignans: Biosynthesis and Function. In:Barton DS, Nakanishi K, Meth-Cohn O editors. Comprehensive Natural Products Chemistry. Pergamon; 1999. https://doi.org/10.1016/B978-0-08-091283-7.00027-8
- 31. Imai T, Nomura M, Fukushima K. Evidence for the involvement of the phenylpropanoid pathway in the biosynthesis of the norlignan agatharesinol. J Plant Physiol. 2006; 163(5):483-7. https://doi.org/10.1016/j. jplph.2005.08.009 PMid:16473652
- 32. Pan J, Chen S, Yang M, Wu J, Jari Sinkkonen, Zou K. An update on lignans: natural products and synthesis. Nat Prod Rep. 2009; 26(10):1251-1. https://doi.org/10.1039/ b910940d PMid:19779640
- 33. Whiting DA. Ligans and Neolignans. Nat Prod Rep. 1985; 2(3):191. https://doi.org/10.1039/np9850200191
- 34. Xu, R., Ye, Y., & Zhao, W. Introduction to Natural Products Chemistry. 1st ed. Boca Raton: CRC Press; 2010. https:// doi.org/10.1201/b11017
- 35. Teponno RB, Kusari S, Spiteller M. Recent advances in research on lignans and neolignans. Nat Prod Rep. 2016; 33(9):1044-92. https://doi.org/10.1039/C6NP00021E PMid:27157413
- 36. Durazzo A. Lignans. In: Nollet, L.M.L., & Gutierrez-Uribe, J.A. editors. Phenolic Compounds in Food: Characterization and Analysis. 1st ed. Boca Raton. CRC Press; 2017. https:// doi.org/10.1201/9781315120157
- 37. Pizzi, A. Advanced Wood Adhesives Technology. 1st ed. Boca Raton: CRC Press; 1994. https://doi. org/10.1201/9781482293548
- 38. Khanbabaee K, Van Ree T. Tannins: classification and definition. Natural product reports. 2001;18(6):641-9. https://doi.org/10.1039/B101061L
- 39. Sharma KP. Tannin degradation by phytopathogen's tannase: A Plant's defence perspective. Biocatal Agric Biotechnol. 2019; 21:101342. https://doi.org/10.1016/j. bcab.2019.101342
- 40. Haslam E. The Metabolism of Gallic Acid and Hexahydroxydiphenic Acid in Higher Plants. In: Herz W., Grisebach H., Kirby G.W editors. Fortschritte der Chemie

<span id="page-10-0"></span>organischer Naturstoffe / Progress in the Chemistry of Organic Natural Products. Vienna. Springer; 1982. https:// doi.org/10.1007/978-3-7091-8656-5\_1

- 41. Hillis WE. Biosynthesis of tannins. In: Higuchi T editors. Biosynthesis and Biodegradation of Wood Components. London. Academic Press; 1985. https://doi.org/10.1016/B978- 0-12-347880-1.50017-9
- 42. Filgueira D, Moldes D, Fuentealba C, García DE. Condensed tannins from pine bark: A novel wood surface modifier assisted by laccase. Ind Crops Prod. 2017; 103:185-94. https://doi.org/10.1016/j.indcrop.2017.03.040
- 43. Pizzi A. Tannins: Major sources, properties and applications. In: Belgacem MN, Gandini A editors. Monomers, Polymers and Composites from Renewable Resources. Netherlands. Elsevier; 2008. https://doi.org/10.1016/B978-0-08-045316- 3.00008-9
- 44. Vinson JA, Su X, Zubik L, Bose P. Phenol antioxidant quantity and quality in foods: fruits. J Agric Food Chem. 2001; 49(11):5315-21. https://doi.org/10.1021/jf0009293 PMid:11714322
- 45. Tomás‐Barberán FA, Clifford MN. Dietary hydroxybenzoic acid derivatives - nature, occurrence and dietary burden. J Sci Food Agric. 2000; 80(7):1024-32. https://doi. org/10.1002/(SICI)1097-0010(20000515)80:7<1024::AID-JSFA567>3.0.CO;2-S
- 46. Sharma N, Tiwari N, Vyas M, Khurana N, Muthuraman A, Utreja P. An overview of therapeutic effects of vanillic acid. Plant Arch. 2020; 20(2):3053-9.
- 47. Kaur J, Gulati M, Singh SK, Kuppusamy G, Kapoor B, Mishra V, *et al.* Discovering the multifaceted role of vanillic acid beyond flavours: Nutraceutical and therapeutic potential. Trends Food Sci Technol. 2022; 122:187-200. https://doi. org/10.1016/j.tifs.2022.02.023
- 48. Juurlink BH, Azouz HJ, Aldalati AM, AlTinawi BM, Ganguly P. Hydroxybenzoic acid isomers and the cardiovascular system. Nutr J. 2014; 13(1). https://doi.org/10.1186/1475- 2891-13-63 PMid:24943896 PMCid: PMC4074389
- 49. Rothwell JA, Perez-Jimenez J, Neveu V, Medina-Remón A, M'hiri N, García-Lobato P, *et al.* Phenol-Explorer 3.0: a major update of the Phenol-Explorer database to incorporate data on the effects of food processing on polyphenol content. Database. 2013; 2013:bat070. https://doi.org/10.1093/ database/bat070 PMid:24103452 PMCid: PMC3792339
- 50. Bhuia MS, Rahaman MM, Islam T, Bappi MH, Sikder MI, Hossain KN, *et al.* Neurobiological effects of gallic acid: current perspectives. Chin Med. 2023; 18(1):27. https://doi. org/10.1186/s13020-023-00735-7 PMid:36918923 PMCid: PMC10015939
- 51. Daglia M, Di Lorenzo A, Nabavi SF, Talas ZS, Nabavi SM. Polyphenols: well beyond the antioxidant capacity: gallic acid and related compounds as neuroprotective agents: you are what you eat! Curr Pharm Biotechnol. 2014;

15(4):362-72. https://doi.org/10.2174/13892010150414082 5120737 PMid:24938889

- 52. Joye IJ. Acids and Bases in Food. In: Melton L, Shahidi F, Varelis P editors. Encyclopedia of Food Chemistry. Netherlands. Academic Press; 2019. https://doi.org/10.1016/ B978-0-08-100596-5.21582-5
- 53. del Olmo A, Calzada J, Nuñez M. Benzoic acid and its derivatives as naturally occurring compounds in foods and as additives: Uses, exposure, and controversy. Crit Rev Food Sci Nutr. 2017; 57(14):3084-103. https://doi.org/10.1080/104 08398.2015.1087964 PMid:26587821
- 54. El-Seedi HR, Taher EA, Sheikh BY, Anjum S, Saeed A, AlAjmi MF, et al. Hydroxycinnamic acids: Natural sources, biosynthesis, possible biological activities, and roles in Islamic medicine. In: Atta-ur-Rahman editor. Studies in Natural Products Chemistry. Elsevier; 2018. https://doi.org/10.1016/ B978-0-444-64068-0.00008-5
- 55. Adisakwattana S. Cinnamic acid and its derivatives: Mechanisms for prevention and management of diabetes and its complications. Nutrients. 2017; 9(2):163. https:// doi.org/10.3390/nu9020163 PMid:28230764 PMCid: PMC5331594
- 56. Guzman J. Natural cinnamic acids, synthetic derivatives and hybrids with antimicrobial activity. Molecules. 2014; 19(12):19292-349. https://doi.org/10.3390/ molecules191219292 PMid:25429559 PMCid: PMC6271800
- 57. Shen N, Wang T, Gan Q, Liu S, Wang L, Jin B. Plant flavonoids: Classification, distribution, biosynthesis, and antioxidant activity. Food Chem. 2022; 383:132531. https:// doi.org/10.1016/j.foodchem.2022.132531 PMid:35413752
- 58. Abed SS, Kiranmayi P, Imran K, Lateef SS. Gas Chromatography-Mass Spectrometry (GC-MS) metabolite profiling of *Citrus limon* (L.) Osbeck juice extract was evaluated for its antimicrobial activity against *Streptococcus* mutans. Cureus. 2023; 15(1):e33585. https://doi. org/10.7759/cureus.33585
- 59. Koushki M, Amiri-Dashatan N, Ahmadi N, Abbaszadeh HA, Rezaei-Tavirani M. Resveratrol: A miraculous natural compound for diseases treatment. Food Sci Nutr. 2018; 6(8):2473-90. https://doi.org/10.1002/fsn3.855 PMid:30510749 PMCid: PMC6261232
- 60. Niaz K, Khan F. Analysis of polyphenolics. In: Silva A S, Nabavi S F, Saeedi M, Nabavi S M editors. Recent Advances in Natural Products Analysis. Netherlands. Elsevier; 2020. https://doi.org/10.1016/B978-0-12-816455-6.00003-2
- 61. Álvarez-Caballero J M, Coy-Barrera E. Lignans. In: Nabavi S M, Silva A S editors. Antioxidants Effects in Health. Netherlands. Elsevier; 2022. https://doi.org/10.1016/B978- 0-12-819096-8.00050-1
- 62. Martinez K B, Mackert J D, McIntosh M K. Polyphenols and Intestinal Health. In: Watson R R editors. Nutrition and Functional Foods for Healthy Aging. United Kingdom.

## <span id="page-11-0"></span>**1906** The Nexus between Polyphenols and Gut Microbiota and Their Interplay in....

Academic Press; 2017. https://doi.org/10.1016/B978-0-12- 805376-8.00018-6.

- 63. Amarowicz R, Janiak M. Hydrolysable Tannins. In: Melton L, Shahidi F, Varelis P editors. Encyclopedia of Food Chemistry. Netherlands. Academic Press; 2019. https://doi. org/10.1016/B978-0-08-100596-5.21771-X
- 64. Jandhyala SM, Talukdar R, Subramanyam C, Vuyyuru H, Sasikala M, Reddy DN. Role of the normal gut microbiota. World J Gastroenterol. 2015; 21(29):8787. https://doi. org/10.3748/wjg.v21.i29.8787 PMid:26269668 PMCid: PMC4528021
- 65. Thursby E, Juge N. Introduction to the human gut microbiota. Biochem J. 2017; 474(11):1823-36. https:// doi.org/10.1042/BCJ20160510 PMid:28512250 PMCid: PMC5433529
- 66. Sumathi S, Suganya K, Swathi K, Sudha B, Sneha S. Current understanding of gut microbiota in tackling COVID-19. Ann Phytomed. 2021; 10Sp-Issue1(COVID-19): S4-S12. https://doi.org/10.21276/ap.covid19.2021.10.1.15
- 67. Wang X, Qi Y, Zheng H. Dietary polyphenol, gut microbiota, and health benefits. Antioxidants. 2022; 11(6):1212. https:// doi.org/10.3390/antiox11061212 PMid:35740109 PMCid: PMC9220293
- 68. Dueñas M, Muñoz-González I, Cueva C, Jiménez-Girón A, Sánchez-Patán F, Santos-Buelga C, *et al.* A survey of modulation of gut microbiota by dietary polyphenols. Biomed Res Int. 2015; e850902. https:// doi.org/10.1155/2015/850902 PMid:25793210 PMCid: PMC4352430
- 69. Rajha HN, Paule A, Aragonès G, Barbosa M, Caddeo C, Debs E, *et al.* Recent advances in research on polyphenols: Effects on microbiota, metabolism, and health. Mol Nutr Food Res. 2021; 66(1):2100670. https://doi.org/10.1002/ mnfr.202100670 PMid:34806294
- 70. Zhao Y, Jiang Q. Roles of the polyphenol-gut microbiota interaction in alleviating colitis and preventing colitisassociated colorectal cancer. Adv Nutr. 2021; 12(2):546-65. https://doi.org/10.1093/advances/nmaa104 PMid:32905583 PMCid: PMC8009754
- 71. Sánchez-Patán F, Cueva C, Monagas M, Walton GE, Gibson M. GR, Quintanilla-López JE, *et al. In vitro* fermentation of a red wine extract by human gut microbiota: Changes in microbial groups and formation of phenolic metabolites. J Agric Food Chem. 2012; 60(9):2136-47. https://doi. org/10.1021/jf2040115 PMid:22313337
- 72. Wu T, Chu X, Cheng Y, Tang S, Zogona D, Pan S, *et al.* Modulation of gut microbiota by *Lactobacillus casei* fermented raspberry juice *in vitro* and *in vivo*. Foods. 2021; 10(12):3055. https://doi.org/10.3390/foods10123055 PMid:34945605 PMCid: PMC8702086
- 73. Zhou L, Wang W, Huang J, Ding Y, Pan Z, Zhao Y, *et al. In vitro* extraction and fermentation of polyphenols from

grape seeds (*Vitis vinifera*) by human intestinal microbiota. Food Funct. 2016; 7(4):1959-67. https://doi.org/10.1039/ C6FO00032K PMid:26980065

- 74. Bialonska D, Ramnani P, Kasimsetty SG, Muntha KR, Gibson GR, Ferreira D. The influence of pomegranate by-product and punicalagins on selected groups of human intestinal microbiota. Int J Food Microbiol. 2010; 140(2-3):175- 82. https://doi.org/10.1016/j.ijfoodmicro.2010.03.038 PMid:20452076
- 75. Wang J, Chen Y, Hu X, Feng F, Cai L, Chen F. Assessing the effects of ginger extract on polyphenol profiles and the subsequent impact on the faecal microbiota by simulating digestion and fermentation *in vitro*. Nutrients. 2020; 12(10):3194. https://doi.org/10.3390/nu12103194 PMid:33086593 PMCid: PMC7650818
- 76. Sost MM, Ahles S, Verhoeven J, Verbruggen S, Stevens Y, Venema K. A citrus fruit extract high in polyphenols beneficially modulates the gut microbiota of healthy human volunteers in a validated *in vitro* model of the colon. Nutrients. 2021; 13(11):3915. https://doi.org/10.3390/ nu13113915 PMid:34836169 PMCid: PMC8619629
- 77. Orso G, Solovyev M, Facchiano S, Evgeniia Tyrikova, Sateriale D, Kashinskaya EN, *et al.* Chestnut shell tannins: Effects on intestinal inflammation and dysbiosis in zebrafish. Animals. 2021; 11(6):1538-8. https://doi.org/10.3390/ ani11061538 PMid:34070355 PMCid: PMC8228309
- 78. Li X, Yang L, Xu M, Qiao G, Li C, Lin L, *et al. Smilax china* L. polyphenols alleviate obesity and inflammation by modulating gut microbiota in high fat/high sucrose diet-fed C57BL/6J mice. J Funct Foods. 2021; 77:104332. https:// doi.org/10.1016/j.jff.2020.104332
- 79. Kafantaris I, Basiliki Kotsampasi, Christodoulou V, Eleana Kokka, Paraskevi Kouka, Zoi Terzopoulou, *et al.* Grape pomace improves the antioxidant capacity and faecal microflora of lambs. J Anim Physiol Anim Nutr. 2017; 101(5). https://doi.org/10.1111/jpn.12569 PMid:27753147
- 80. Zhao L, Zhang Q, Ma W, Tian F, Shen H, Zhou M. A combination of quercetin and resveratrol reduces obesity in high-fat diet-fed rats by modulation of gut microbiota. Food Funct. 2017; 8(12):4644-56. https://doi.org/10.1039/ C7FO01383C PMid:29152632
- 81. Cladis DP, Abigayle M.R. Simpson, Cooper KJ, Nakatsu CH, Ferruzzi MG, Weaver CM. Blueberry polyphenols alter gut microbiota and phenolic metabolism in rats. Food Funct. 2021; 12(6):2442-56. https://doi.org/10.1039/D0FO03457F PMid:33629093 PMCid: PMC8011555
- 82. Molan AL, Liu Z, Plimmer G. Evaluation of the effect of blackcurrant products on gut microbiota and on markers of risk for colon cancer in humans. Phytother Res. 2014; 28(3):416- 22. https://doi.org/10.1002/ptr.5009 PMid:23674271
- 83. Tzounis X, Rodriguez-Mateos A, Vulevic J, Gibson GR, Kwik-Uribe C, Spencer JP. Prebiotic evaluation of cocoa-derived

<span id="page-12-0"></span>flavanols in healthy humans by using a randomized, controlled, double-blind, crossover intervention study. Am J Clin Nutr. 2011; 93(1):62-72. https://doi.org/10.3945/ ajcn.110.000075 PMid:21068351

- 84. Espín JC, González-Sarrías A, Tomás-Barberán FA. The gut microbiota: A key factor in the therapeutic effects of (poly) phenols. Biochem Pharmacol. 2017; 139:82-93. https://doi. org/10.1016/j.bcp.2017.04.033 PMid:28483461
- 85. Shah SR, Alweis R, Najim NI, Dharani AM, Jangda MA, Shahid M, Kazi AN, Shah SA. Use of dark chocolate for diabetic patients: A review of the literature and current evidence. J Community Hosp Intern Med Perspect. 2017; 7(4):218-21. https://doi.org/10.1080/20009666.2017.13612 93 PMid:29181133 PMCid: PMC5699188
- 86. Johnston K, Sharp P, Clifford M, Morgan L. Dietary polyphenols decrease glucose uptake by human intestinal Caco-2 cells. FEBS Lett. 2005; 579(7):1653-7. https://doi. org/10.1016/j.febslet.2004.12.099 PMid:15757656
- 87. Pandurangan AK, Mohebali N, Mohd. Esa N, Looi CY, Ismail S, Saadatdoust Z. Gallic acid suppresses inflammation in dextran sodium sulfate-induced colitis in mice: Possible mechanisms. Int Immunopharmacol. 2015; 28(2):1034- 43. https://doi.org/10.1016/j.intimp.2015.08.019 PMid:26319951
- 88. Yang K, Zhang L, Liao P, Xiao Z, Zhang F, Sindaye D, *et al.* Impact of gallic acid on gut health: Focus on the gut microbiome, immune response, and mechanisms of action. Front Immunol. 2020; 11. https://doi.org/10.3389/ fimmu.2020.580208 PMid:33042163 PMCid: PMC7525003
- 89. Li Y, Xie Z, Gao T, Li L, Chen Y, Xiao D, *et al.* A holistic view of gallic acid-induced attenuation in colitis based on microbiome-metabolomics analysis. Food Funct. 2019; 10(7):4046-61. https://doi.org/10.1039/C9FO00213H PMid:31225554
- 90. Coelho LP, Kultima JR, Costea PI, Fournier C, Pan Y, Czarnecki-Maulden G, *et al.* Similarity of the dog and human gut microbiomes in gene content and response to diet. Microbiome. 2018; 6(1). https://doi.org/10.1186/ s40168-018-0450-3 PMid:29669589 PMCid: PMC5907387
- 91. Yang K, Deng X, Jian S, Zhang M, Wen C, Xin Z, *et al.* Gallic acid alleviates gut dysfunction and boosts immune and antioxidant activities in puppies under environmental stress based on microbiome-metabolomics analysis. Front Immunol. 2022; 12. https://doi.org/10.3389/ fimmu.2021.813890 PMid:35095912 PMCid: PMC8795593
- 92. Lima ACD, Cecatti C, Fidélix MP, Adorno MAT, Sakamoto IK, Cesar TB, *et al.* Effect of daily consumption of orange juice on the levels of blood glucose, lipids, and gut microbiota metabolites: Controlled clinical trials. J Med Food. 2019; 22(2):202-10. https://doi.org/10.1089/ jmf.2018.0080 PMid:30638420
- 93. Surai PF. Polyphenol compounds in the chicken/animal diet: from the past to the future. J Anim Physiol Anim

Nutr. 2014; 98(1):19-31. https://doi.org/10.1111/jpn.12070 PMid:23527581

- 94. Zhang C, Zhao XH, Yang L, Chen XY, Jiang RS, Jin SH, *et al.* Resveratrol alleviates heat stress-induced impairment of intestinal morphology, microflora, and barrier integrity in broilers. Poult Sci. 2017; 96(12):4325-32. https://doi. org/10.3382/ps/pex266 PMid:29053872
- 95. Khampeerathuch T, Mudsak A, Srikok S, Vannamahaxay S, Chotinun S, Chuammitri P. Differential gene expression in heterophils isolated from commercial hybrid and Thai indigenous broiler chickens under quercetin supplementation. J Appl Anim Res. 2018; 46(1):804-12. https://doi.org/10.1080/09712119.2017.1405814
- 96. Mattio LM, Dallavalle S, Musso L, Filardi R, Franzetti L, Pellegrino L, *et al*. Antimicrobial activity of resveratrolderived monomers and dimers against foodborne pathogens. Sci Rep. 2019; 9(1). https://doi.org/10.1038/ s41598-019-55975-1 PMid:31862939 PMCid: PMC6925292
- 97. Duda-Chodak A, Tarko T, Satora P, Sroka P. Interaction of dietary compounds, especially polyphenols, with the intestinal microbiota: A review. Eur J Nutr. 2015; 54(3):325-41. https://doi.org/10.1007/s00394-015-0852-y PMid:25672526 PMCid: PMC4365176
- 98. Gu Y, Zhang Y, Li M, Huang Z, Jiang J, Chen Y, *et al.* Ferulic acid ameliorates atherosclerotic injury by modulating gut microbiota and lipid metabolism. Front Pharmacol. 2021; 12. https://doi.org/10.3389/fphar.2021.621339 PMid:33841148 PMCid: PMC8026864
- 99. Guo J, Han X, Zhan J, You Y, Huang W. Vanillin alleviates high-fat diet-induced obesity and improves the gut microbiota composition. Front Microbiol. 2018; 9. https://doi.org/10.3389/ fmicb.2018.02733 PMid:30483238 PMCid: PMC6243071
- 100. Jimenez-Garcia SN, Guevara-Gonzalez RG, Miranda-Lopez R, Feregrino-Perez AA, Torres-Pacheco I, Vazquez-Cruz MA. Functional properties and quality characteristics of bioactive compounds in berries: Biochemistry, biotechnology, and genomics. Food Res Int. 2013; 54(1):1195-207. https://doi.org/10.1016/j. foodres.2012.11.004
- 101. Vendrame S, Del Bo' C, Ciappellano S, Riso P, Klimis-Zacas D. Berry fruit consumption and metabolic syndrome. Antioxidants. 2016; 5(4):34. https://doi.org/10.3390/ antiox5040034 PMid:27706020 PMCid: PMC5187532
- 102. Bouyahya A, Omari NE, El Hachlafi N, Jemly ME, Hakkour M, Balahbib A, *et al*. Chemical compounds of berry-derived polyphenols and their effects on gut microbiota, inflammation, and cancer. Molecules (Basel, Switzerland). 2022; 27(10):3286. https://doi.org/10.3390/ molecules27103286 PMid:35630763 PMCid: PMC9146061
- 103. Xiang Q, Liu Y, Wu Z, Wang R, Zhang X. New hints for improving sleep: Tea polyphenols mediate gut microbiota to regulate circadian disturbances. Food Front. 2023; 4(1):47- 59. https://doi.org/10.1002/fft2.199

## <span id="page-13-0"></span>**1908** The Nexus between Polyphenols and Gut Microbiota and Their Interplay in....

- 104. Yan R, Ho CT, Zhang X. Modulatory effects in circadianrelated diseases via the reciprocity of tea polyphenols and intestinal microbiota. Food Sci Hum Wellness. 2022; 11(3):494-501. https://doi.org/10.1016/j.fshw.2021.12.007
- 105. Zhang Y, Cheng L, Liu Y, Wu Z, Weng P. The intestinal microbiota links tea polyphenols with the regulation of mood and sleep to improve immunity. Food Reviews International. 2023; 39(3):1485-98. https://doi.org/10.1080 /87559129.2021.1934007
- 106. Xie Y, Wu Z, Qian Q, Yang H, Ma J, Luan W, Shang S, Li X. Apple polyphenol extract ameliorates sugary-dietinduced depression-like behaviours in male C57BL/6 mice by inhibiting the inflammation of the gut-brain axis. Food Funct. 2024; 15(6):2939-59. https://doi.org/10.1039/ D3FO04606K PMid:38406886
- 107. Zhou P, Feng P, Liao B, Fu L, Shan H, Cao C, *et al.* Role of polyphenols in remodelling the host gut microbiota in polycystic ovary syndrome. J Ovarian Res. 2024;

17(1):69. https://doi.org/10.1186/s13048-024-01354-y PMid:38539230 PMCid: PMC10967125

- 108. Dong L, Qin C, Li Y, Wu Z, Liu L. Oat phenolic compounds regulate metabolic syndrome in high fat diet-fed mice via gut microbiota. Food Biosci. 2022; 50:101946. https://doi. org/10.1016/j.fbio.2022.101946
- 109. Huang G, Khan R, Zheng Y, Lee PC, Li Q, Khan I. Exploring the role of gut microbiota in advancing personalized medicine. Front Microbiol. 2023; 14:1274925. https://doi. org/10.3389/fmicb.2023.1274925 PMid:38098666 PMCid: PMC10720646
- 110. Koppel N, Maini Rekdal V, Balskus EP. Chemical transformation of xenobiotics by the human gut microbiota. Science. 2017; 356(6344):eaag2770. https://doi.org/10.1126/ science.aag2770 PMid:28642381 PMCid: PMC5534341
- 111. Ryu G, Kim H, Koh A. Approaching precision medicine by tailoring the microbiota. Mamm Genome. 2021; 32:206-22. https://doi.org/10.1007/s00335-021-09859-3 PMid:33646347